

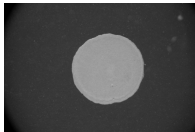
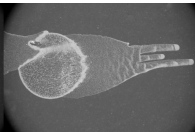
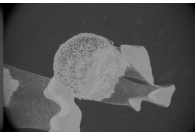

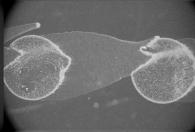
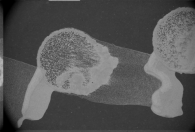
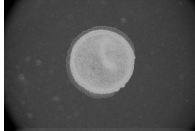
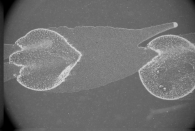
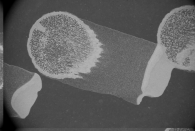
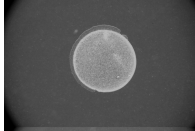
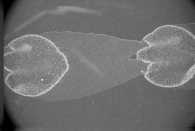
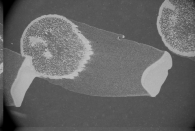

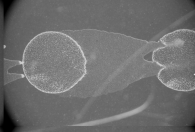
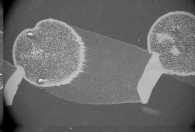
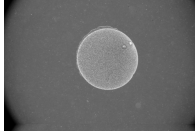
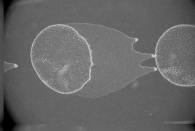
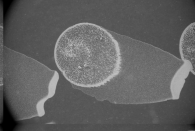
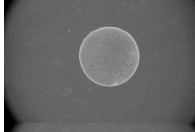
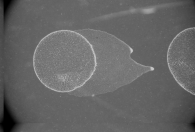
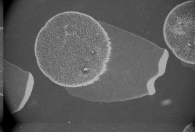
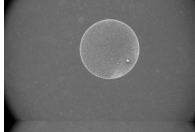

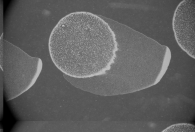
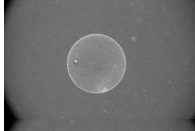
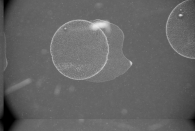
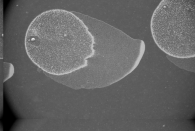
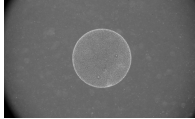
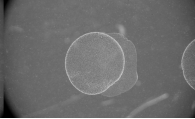
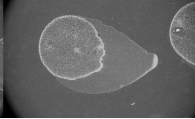
wavelengths

1Aug2013

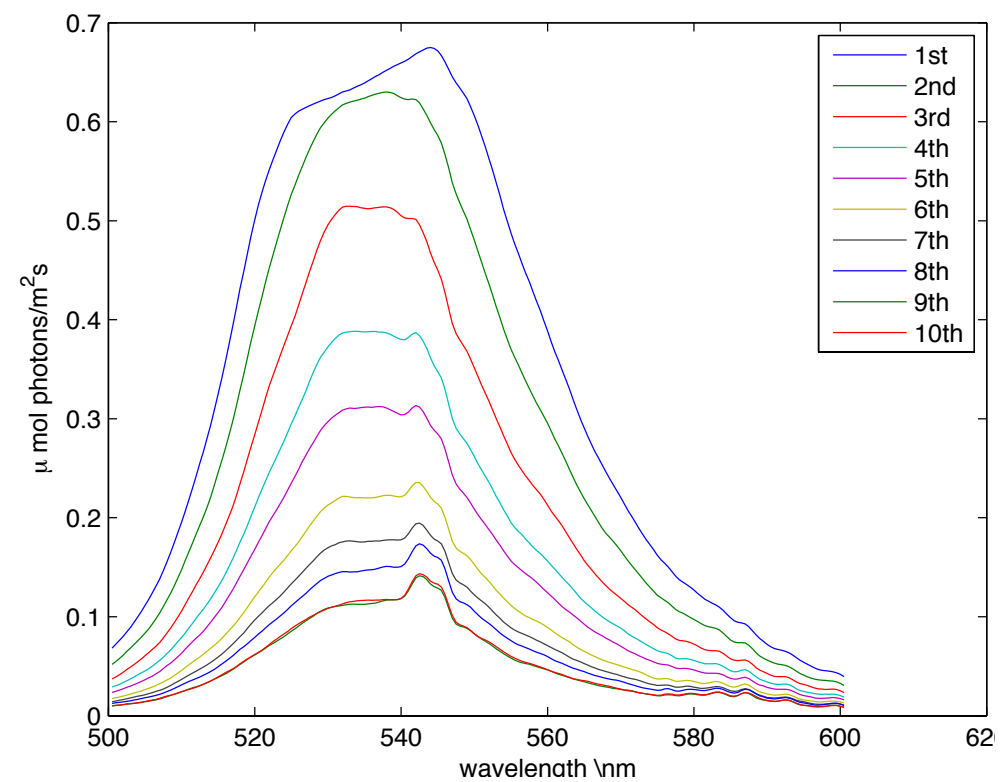
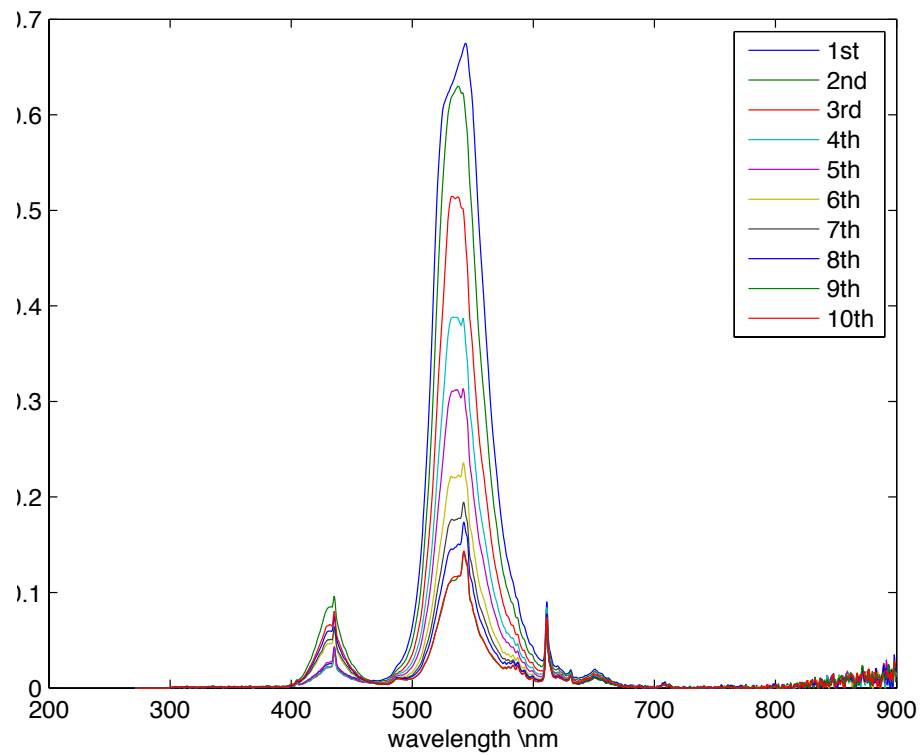
Day2

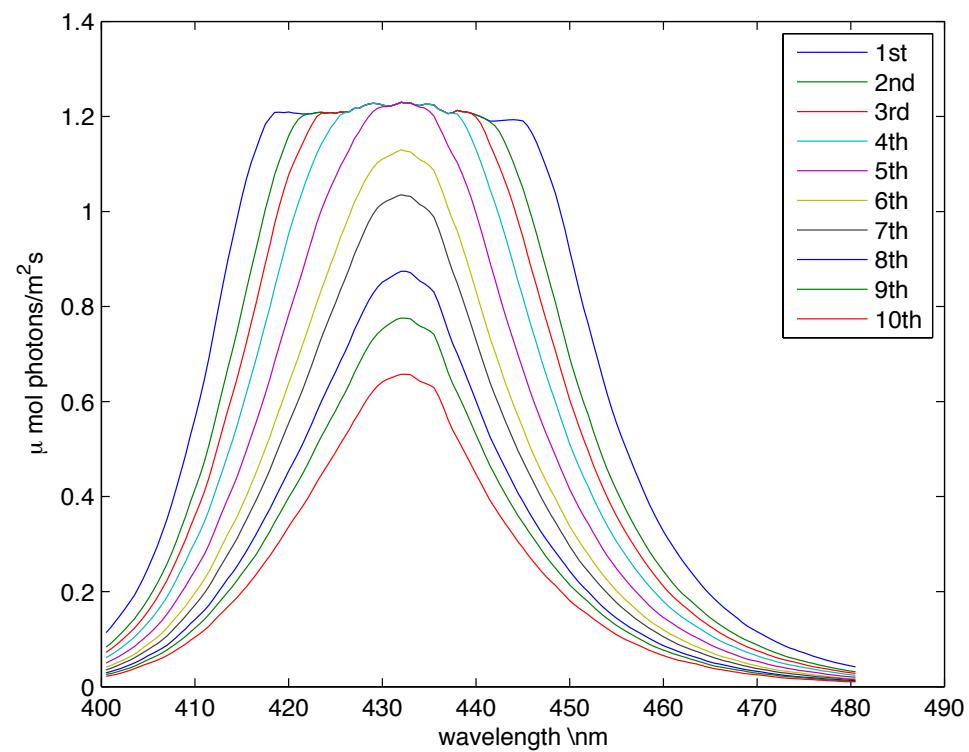
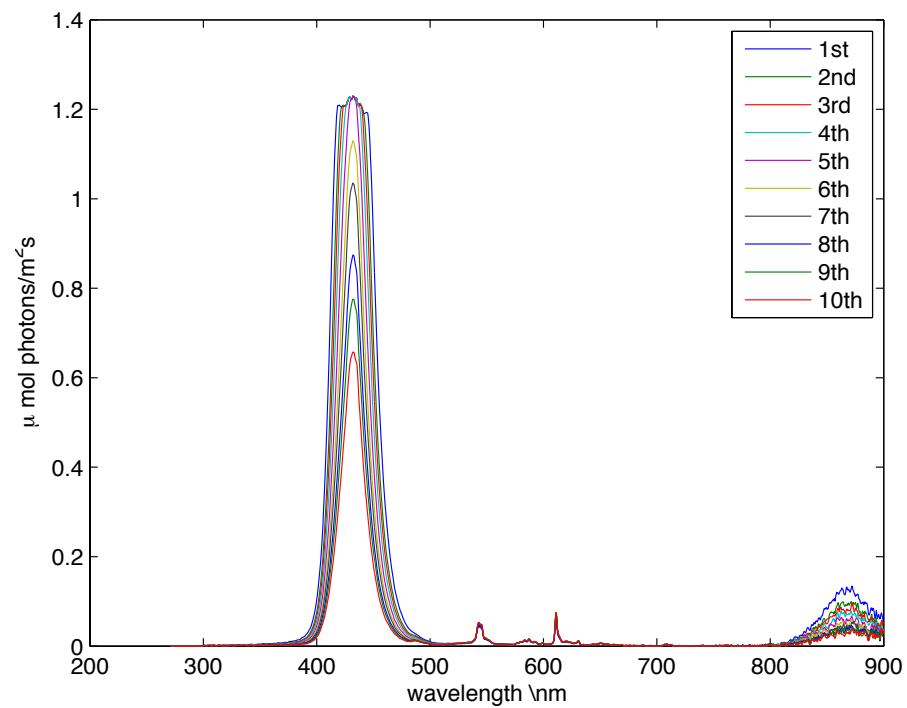
For the intensities tested,  
535nm and 660nm show an  
intensity-dependent  
response. 435nm shows some  
forward movement after 3  
days at the higher intensities.

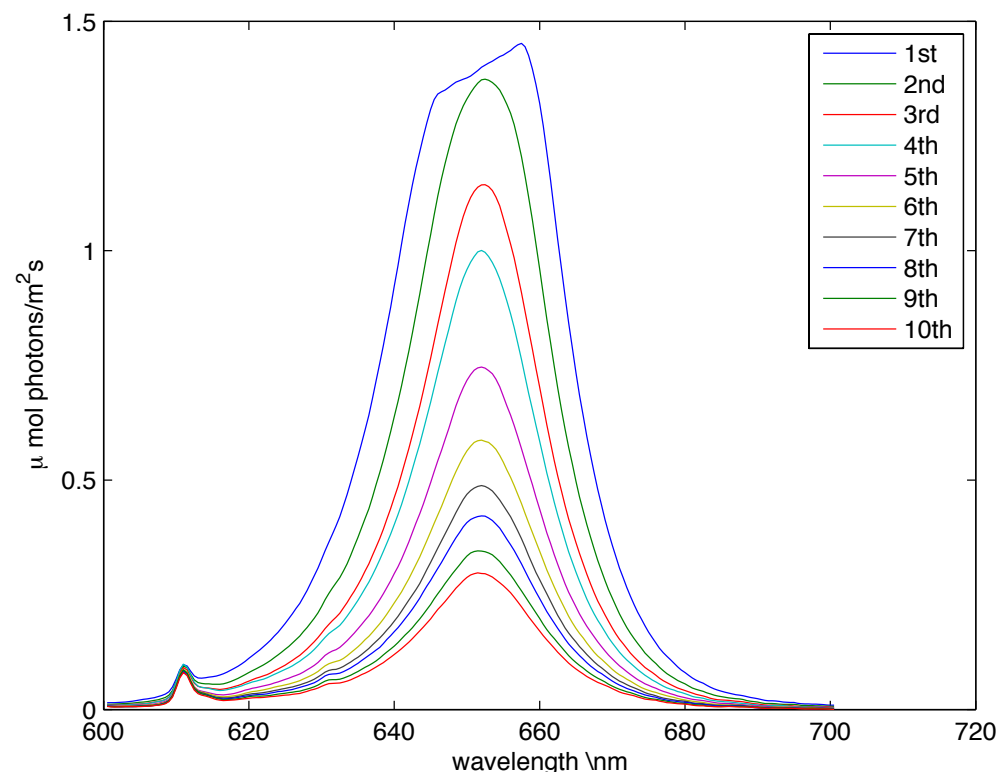
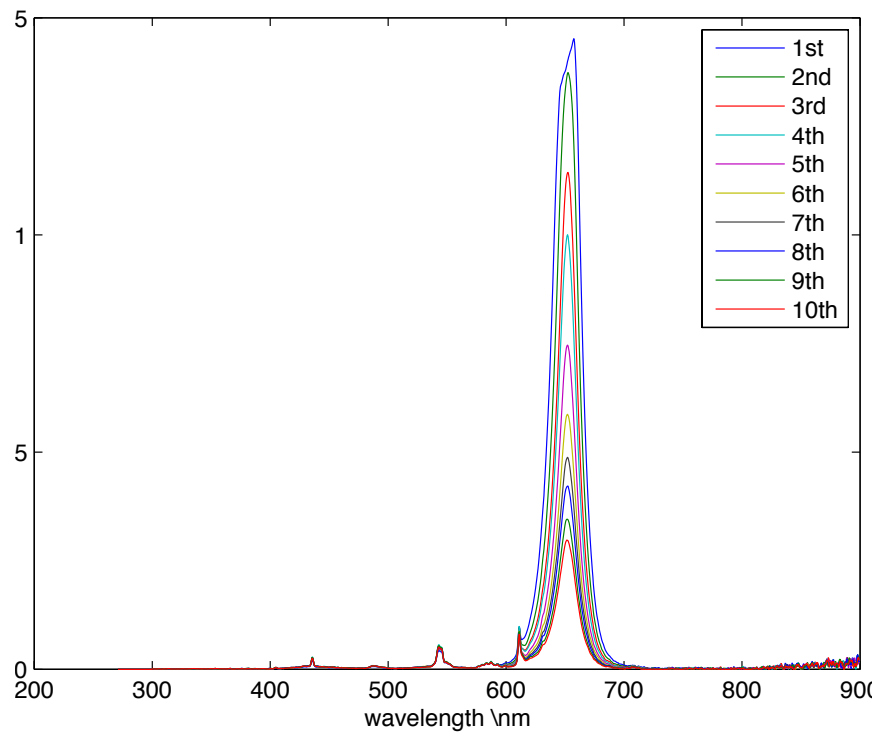
Done using  
square plate

	435nm	535nm	660nm		
144				42	45
115				32	34
90				24	29
75				20	20
58				15	18
50				14	16
42				12	14
35				11	12
30				10	11
24				9	9









7Aug2013

435nm 535nm 660nm

For the intensities tested, 535nm and 660nm show an intensity-dependent response. 435nm shows some forward movement after 3 days at the higher intensities.

Day2

144

115

90

75

58

50

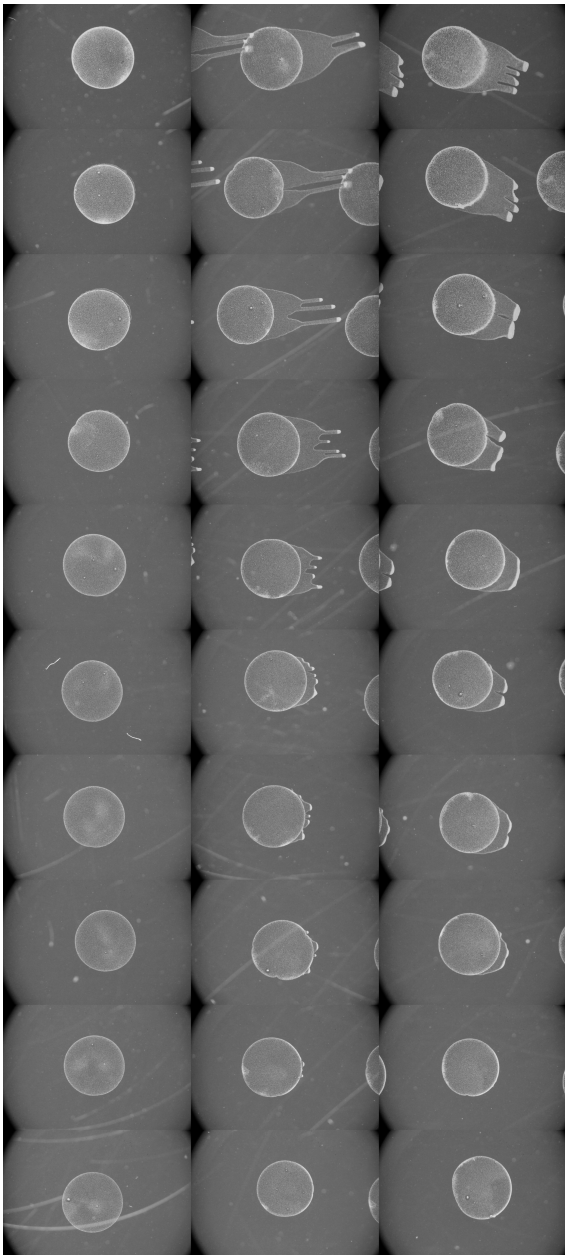
42

35

30

24

Done using square plate



42 45

32 34

24 29

20 20

15 18

14 16

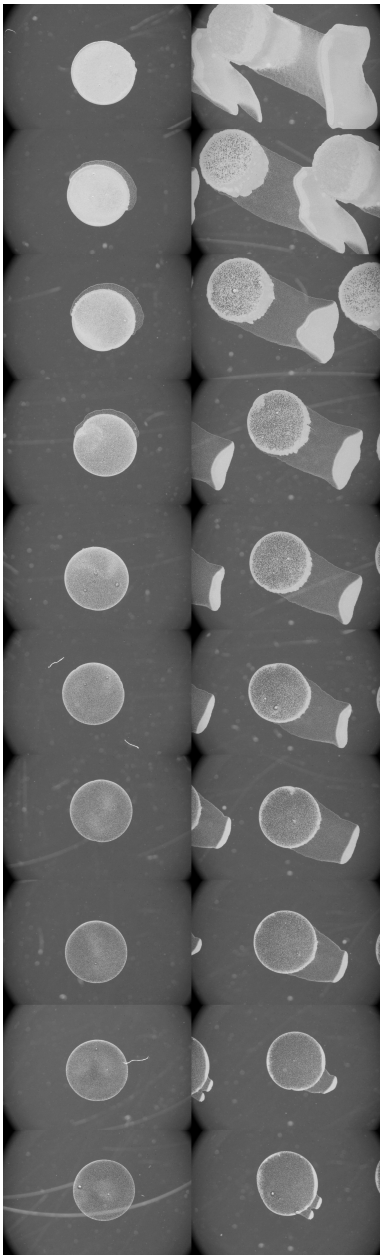
12 14

11 12

10 11

9 9

Day4



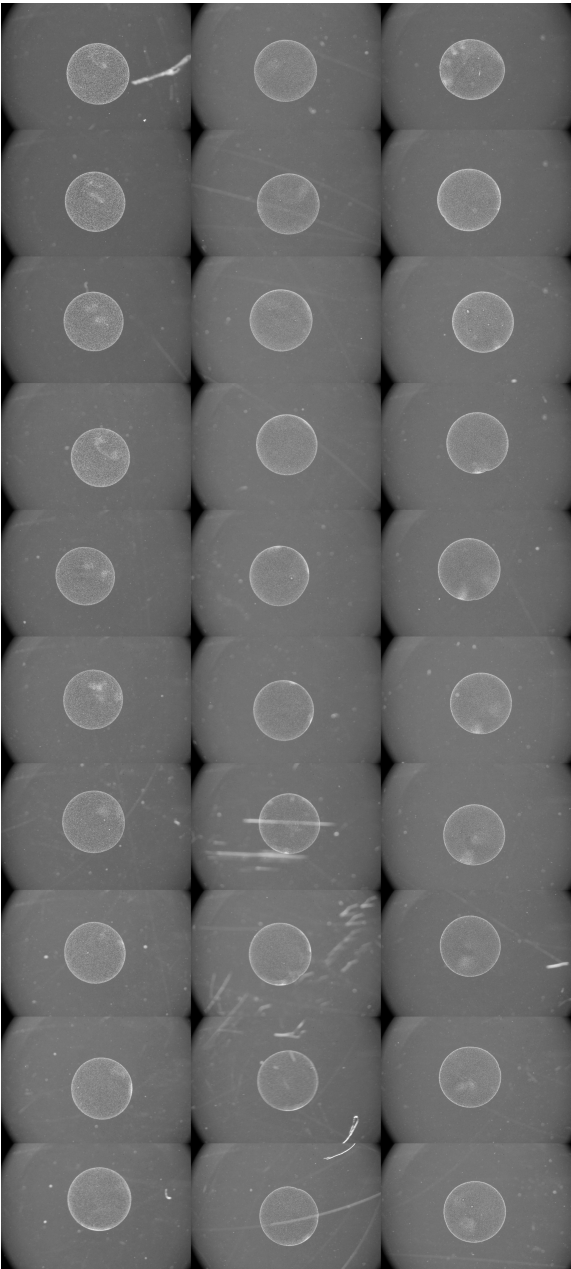
- At high 435nm intensities, the drop extends a single wide front, if any, toward light.
  - From what we know about TaxD1, the active blue-absorbing form converts to the inactive green-absorbing form after absorbing blue light.
  - From what we know from the taxD1 mutant, TaxD1 seems to suppress sideways noise in motility. Are we seeing the same effect?
  - But if TaxD1 is being inactivated, why are the cells not moving backward?
  - PixD (BLUF) also absorbs at 440nm, so are we seeing the effect of PixD as well? (pixD mutant is a backmover under 660nm)
- Under 535nm, the cells exhibit an intensity-dependent response
  - Is this a demonstration of TaxD1 converting to a higher ratio of the active blue-absorbing form?
  - UIRs absorb at 534nm, and the mutant is a forward mover under blue and green.
- Next steps: taxD1 mutant under the same intensities, to see if we are seeing the effect of inactivating TaxD1 under high blue intensities, and activating under high green.

13Aug2013

Day2

435nm 535nm 435nm

144



22.5 85

115

17 63

90

13.5 50

75

11 38

58

10 30

50

8.5 23

42

8 20

35

7 16

30

6.5 14

24

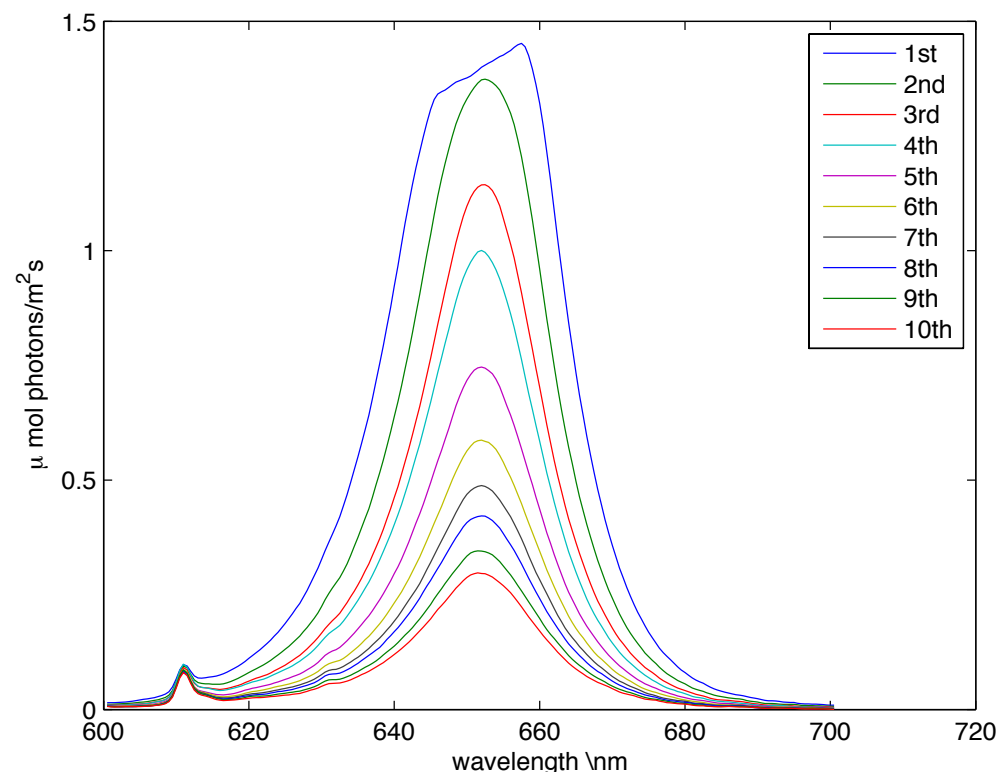
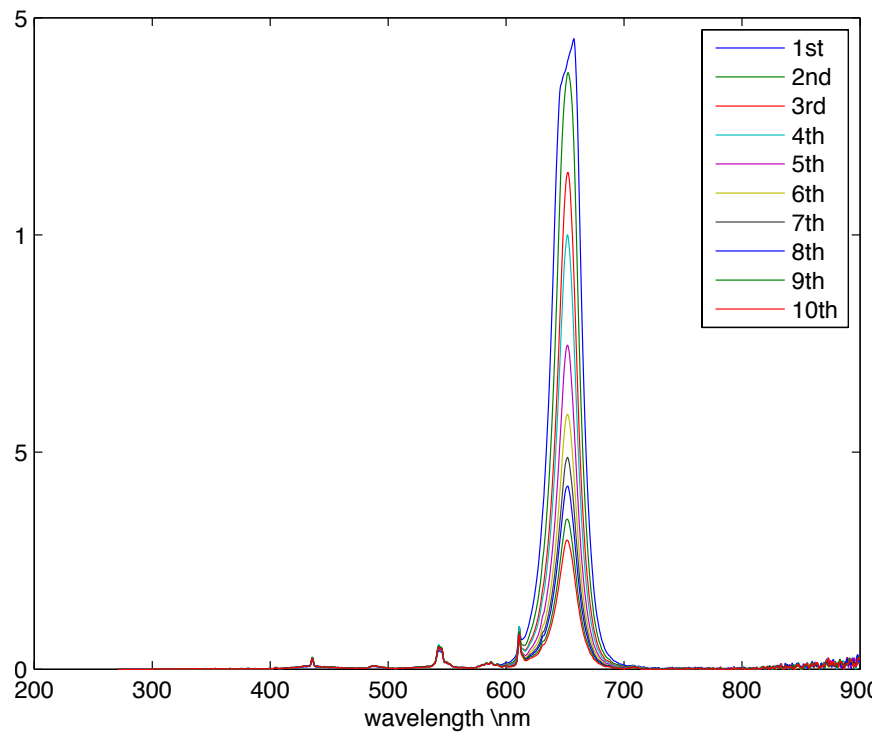
6 12

Testing low  
535nm and  
435nm  
intensities

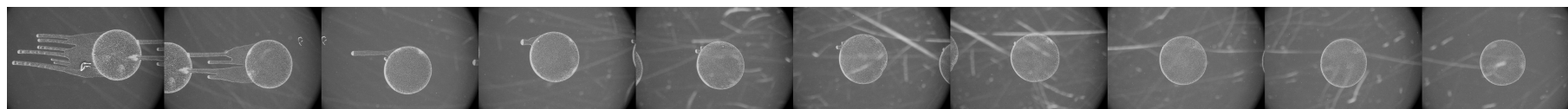
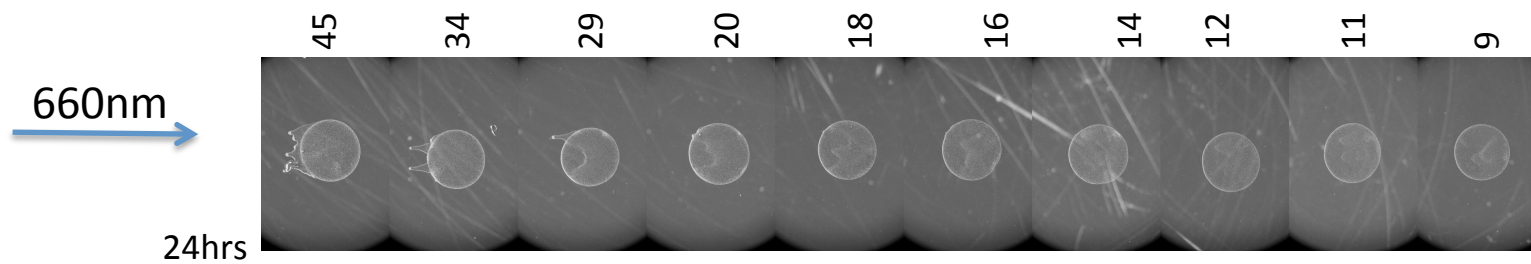
Done using  
square plate

## Single-cell microscopy at early timepoints

- To determine if the different observed fingering patterns are the result of intensity-dependent phototactic responses (and not higher cell density due to division at increasing intensities)
- 10-min videos taken every 2 hours, starting at 0-hr after light-on
  - At 0, 2, 4, 6, 8, 24 hrs
- 4 drops were imaged in each experiment

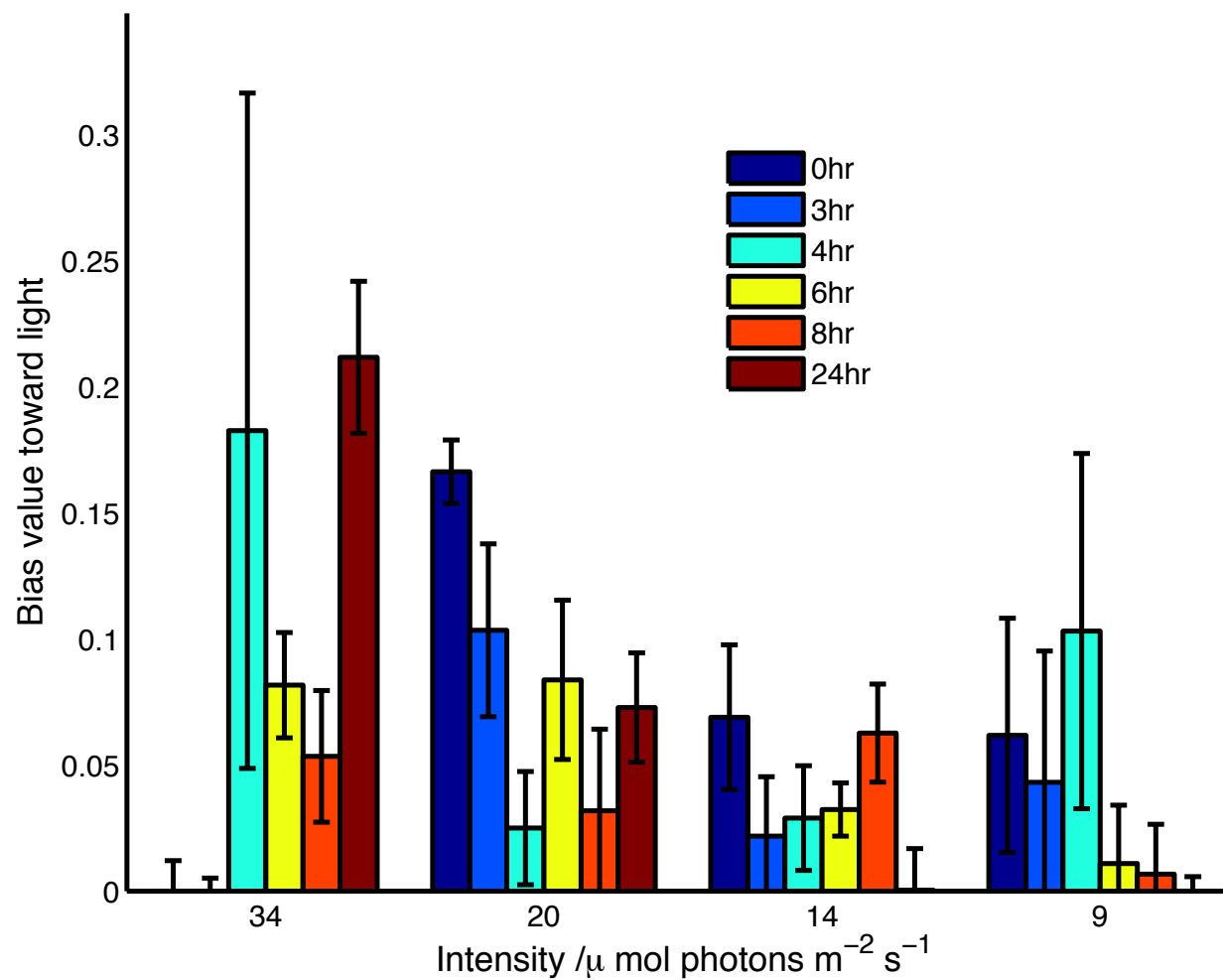


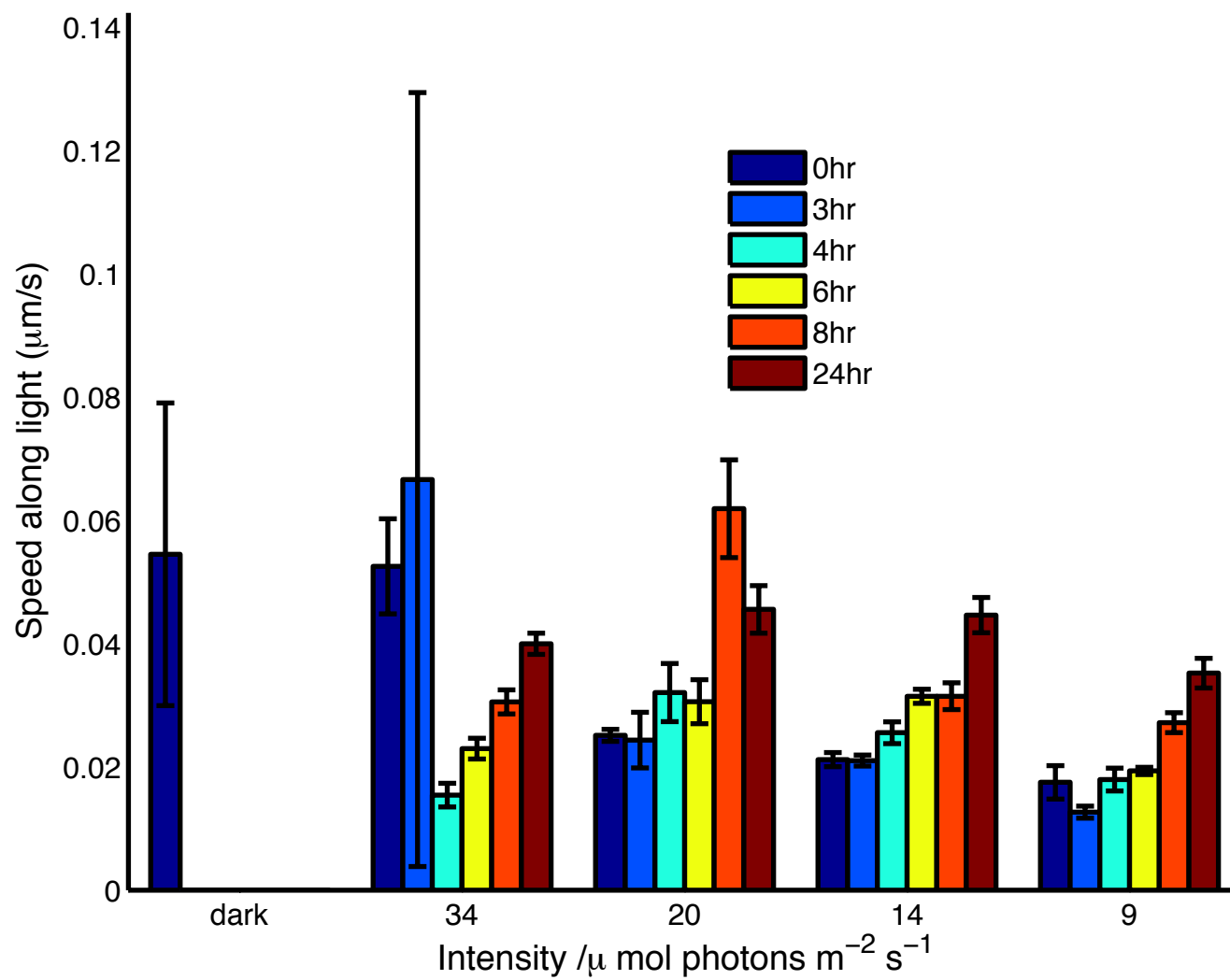




48hrs

Plotting the mean bias value across 6 100-sec intervals in each case

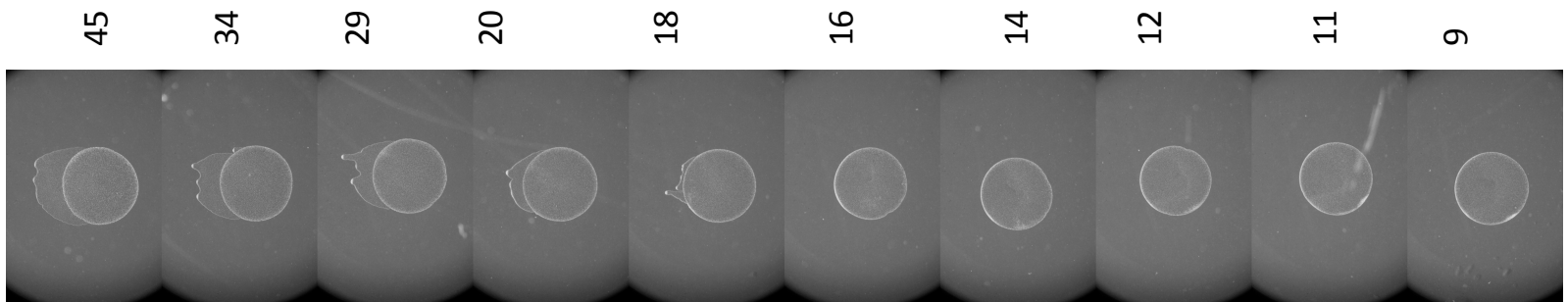




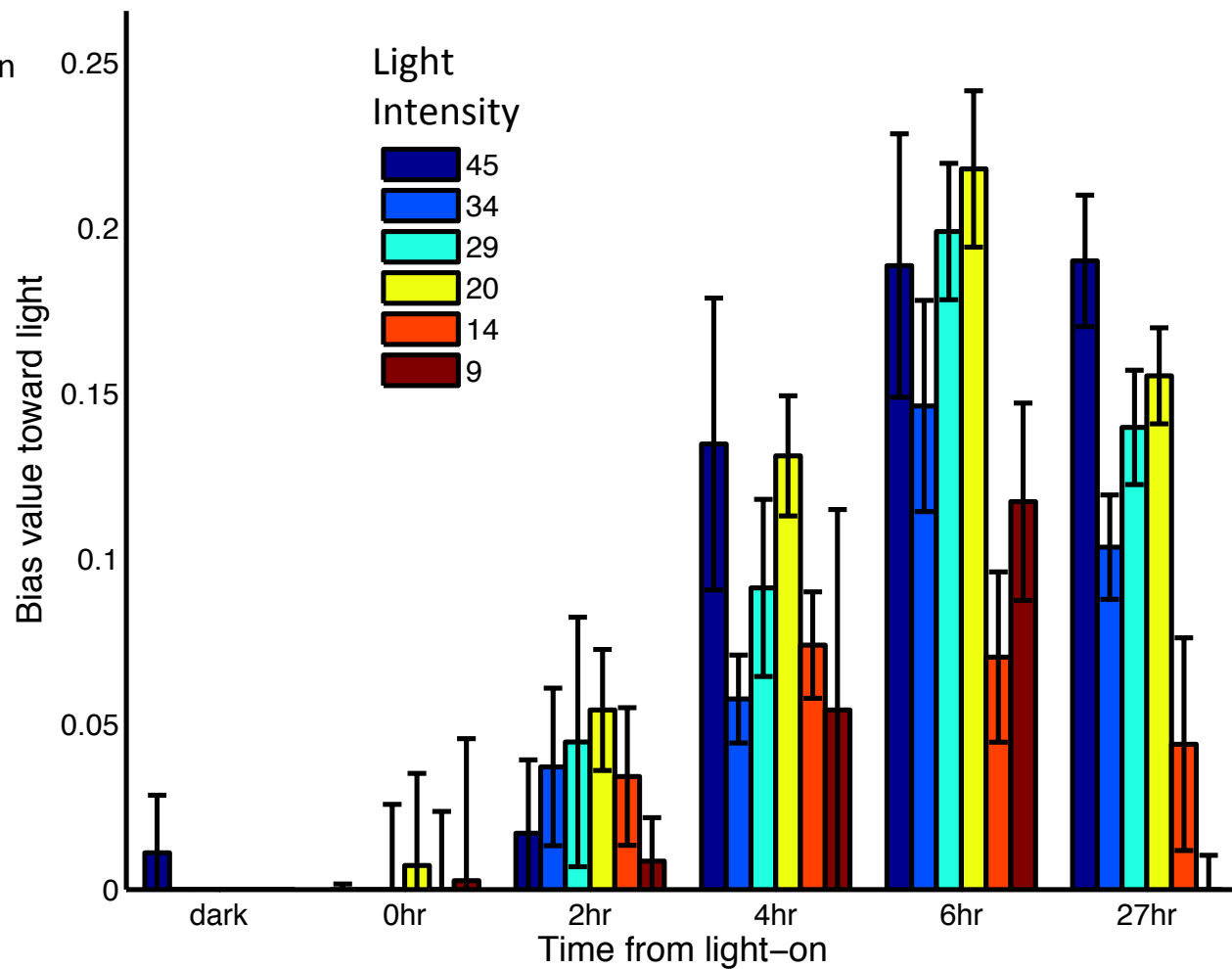
19Sept  
experiment

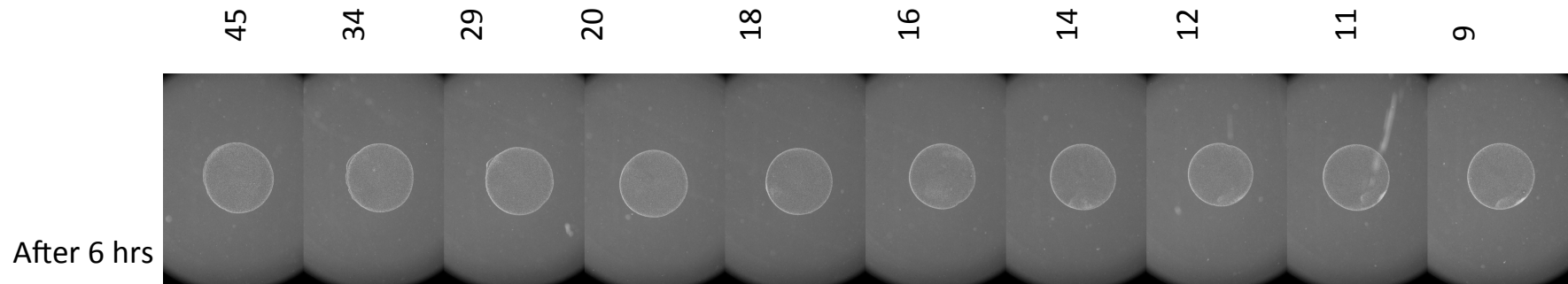
660nm  
→

After 27 hrs

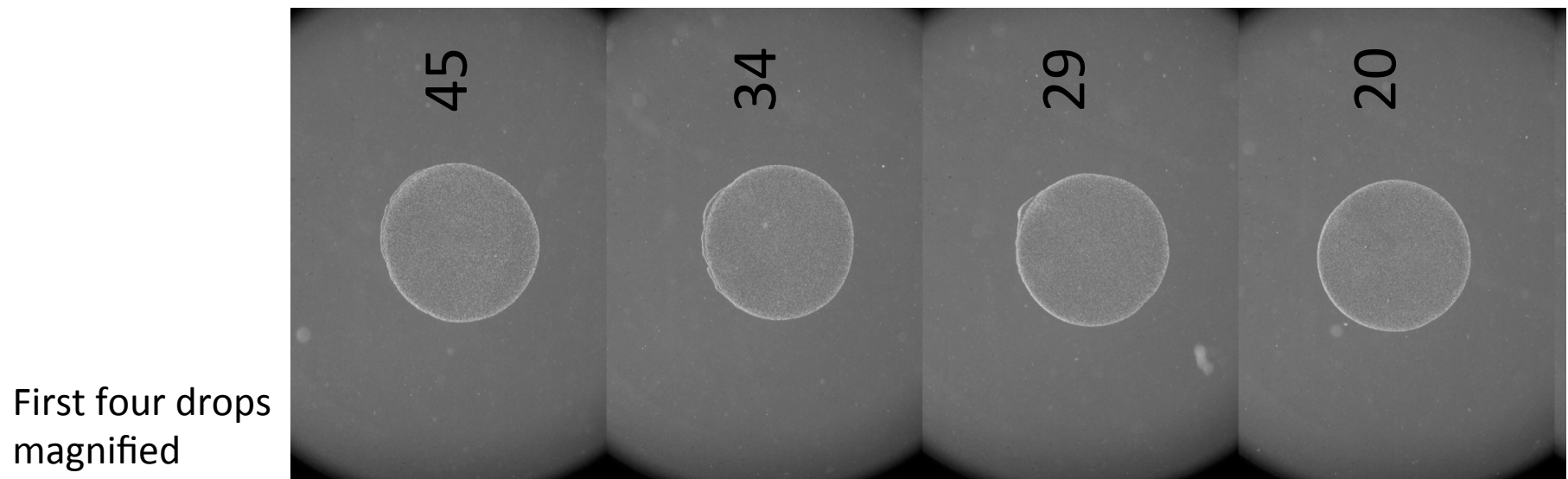


In this expt, I  
measured the bias in  
6 drops.

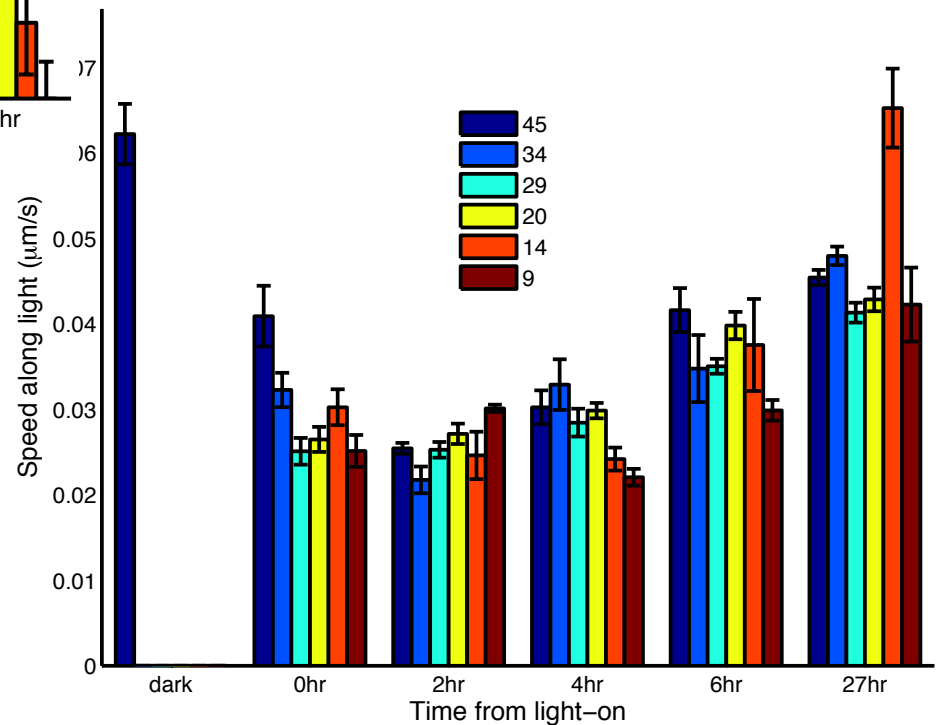
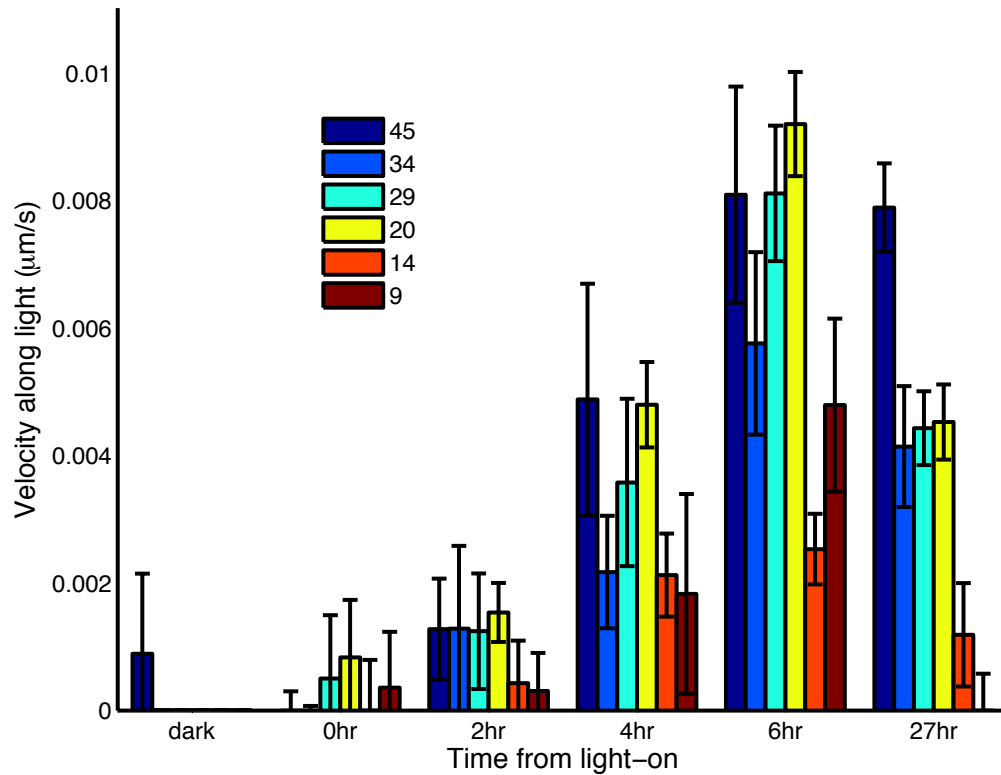




- I was surprised that there wasn't a clear trend in the bias values at 6-hrs, esp. from the drops experiencing 45, 34, 29, 20  $\mu\text{E}$ , because cells had already clearly moved out of the respective drops by then.

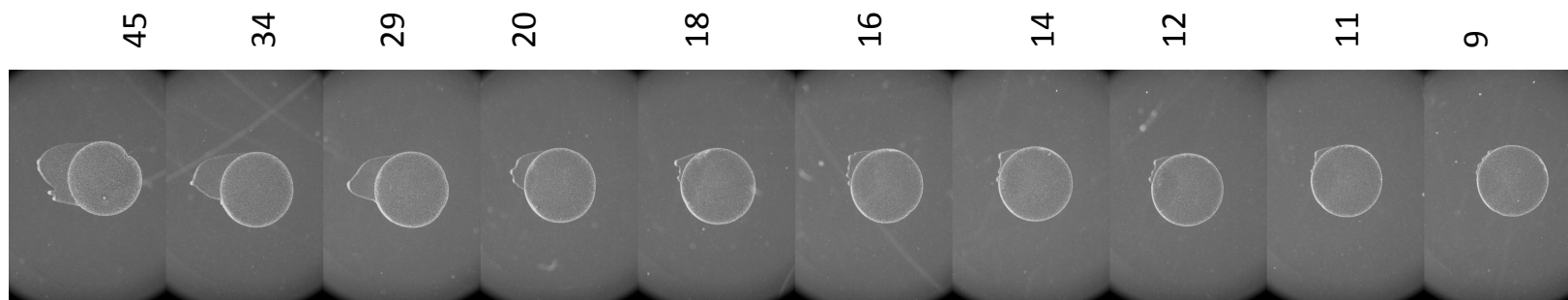


Couldn't see a good trend in the velocities or speeds either

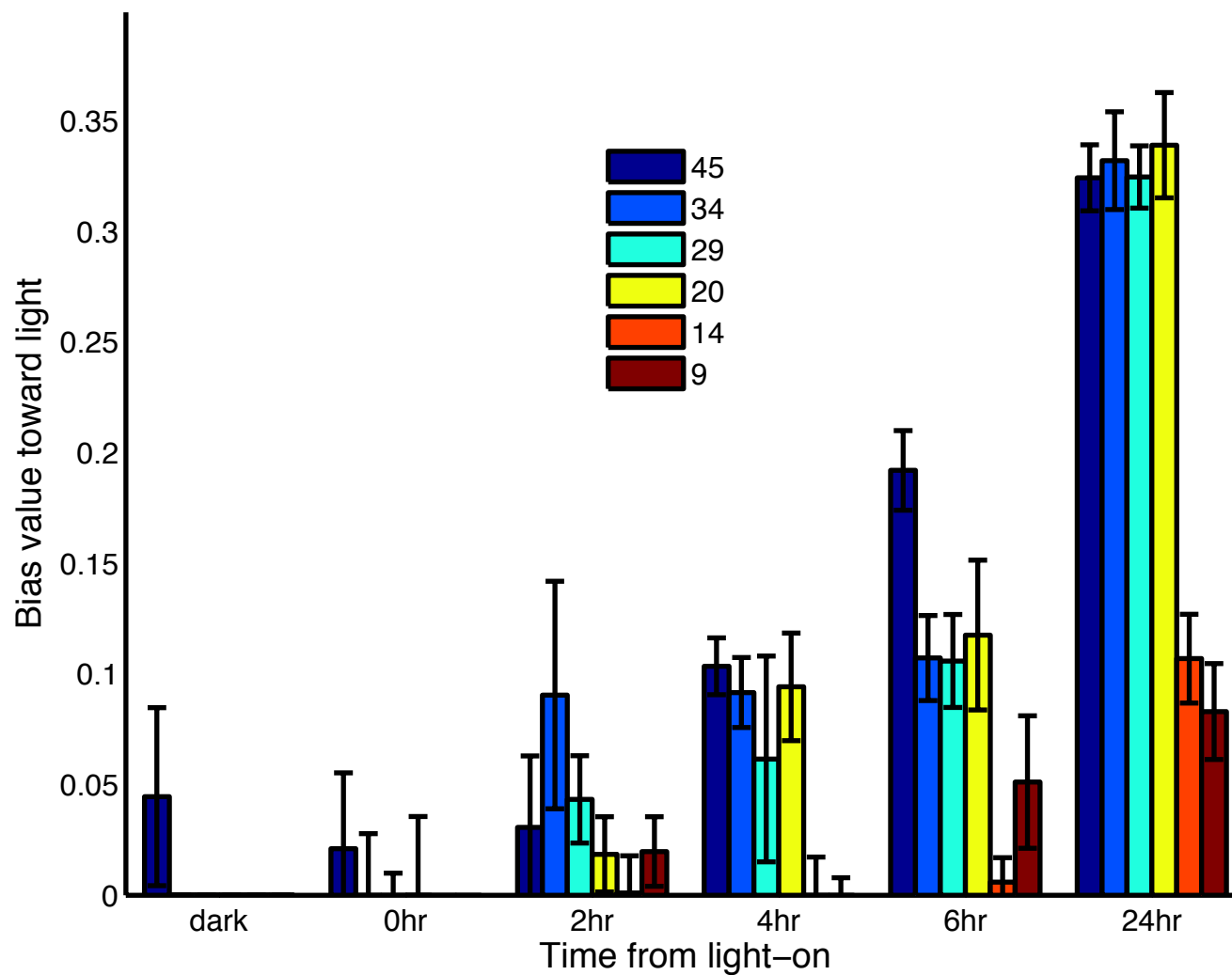


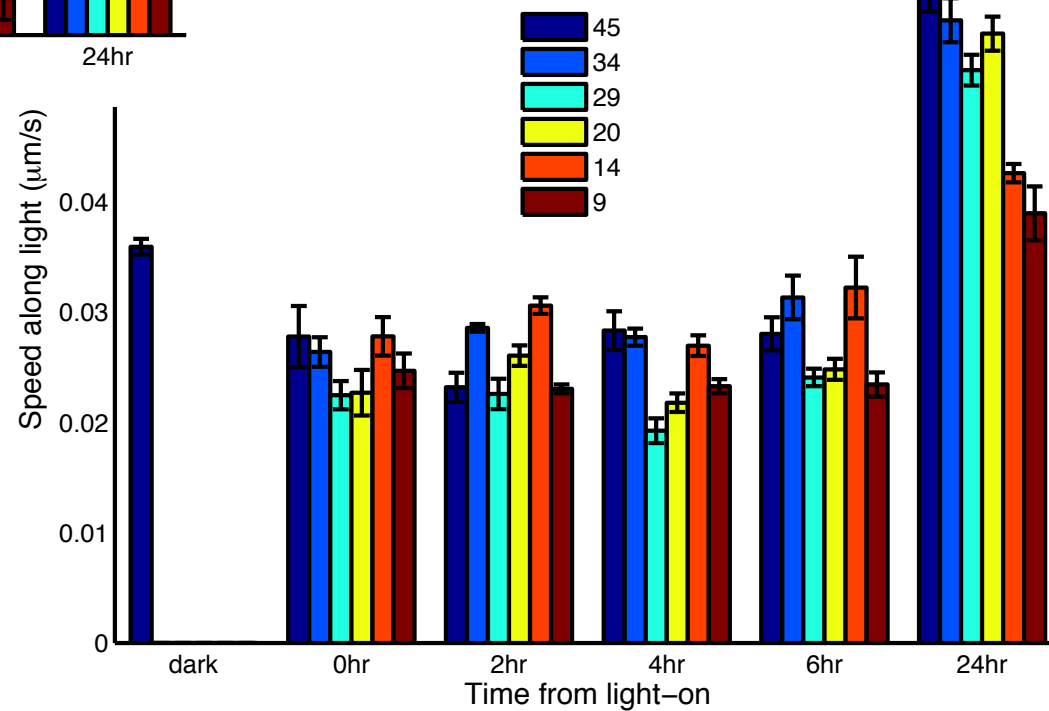
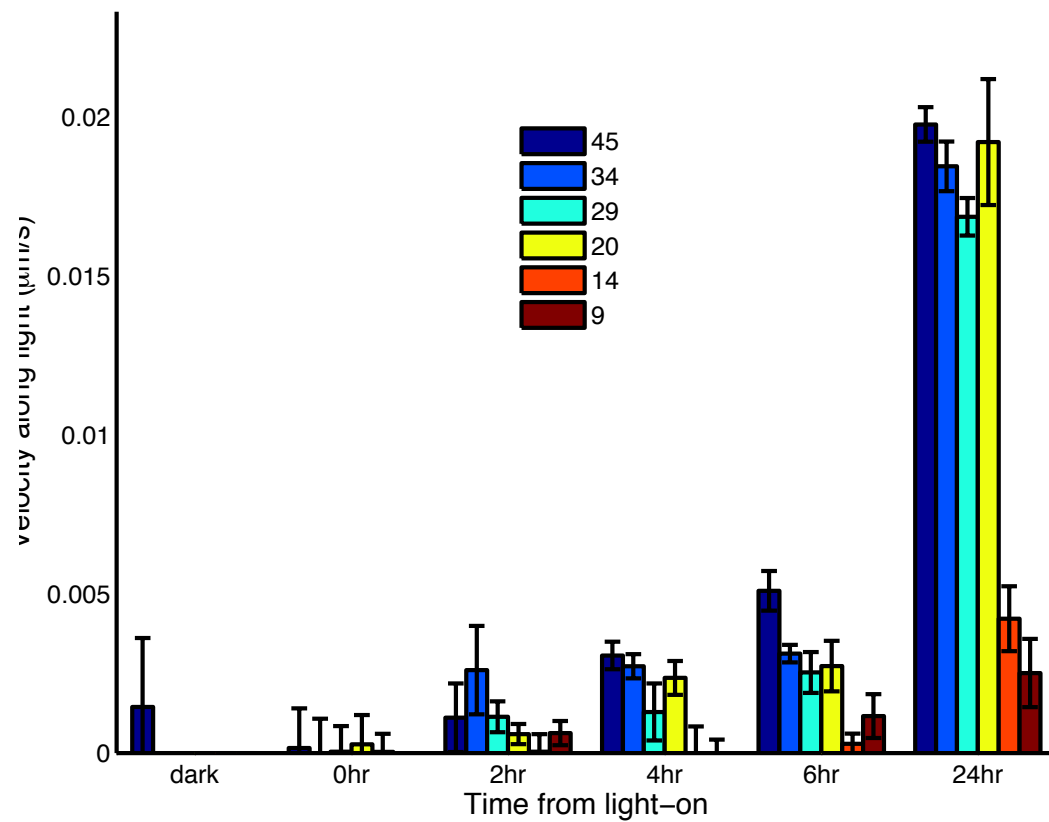
So I tried to repeat the experiment, to see if I can get the same quick finger formation and image the cells in the initial fingers..

27Sept  
expt



But the cells didn't  
move out of the  
drop till many hrs  
after, so I was only  
able to image the  
front of the drop



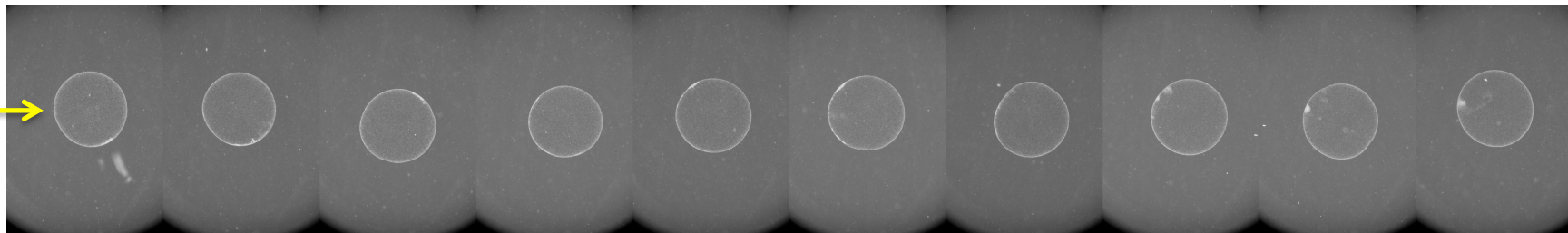


- Experiment done on 20 Nov
- 660nm tested at 10 intensities
  - 45, 34, 29, 20, 18, 16, 14, 12, 11, 9
- Cells were imaged at the front of each drop every 6 hrs, skipping the 18<sup>th</sup> hour

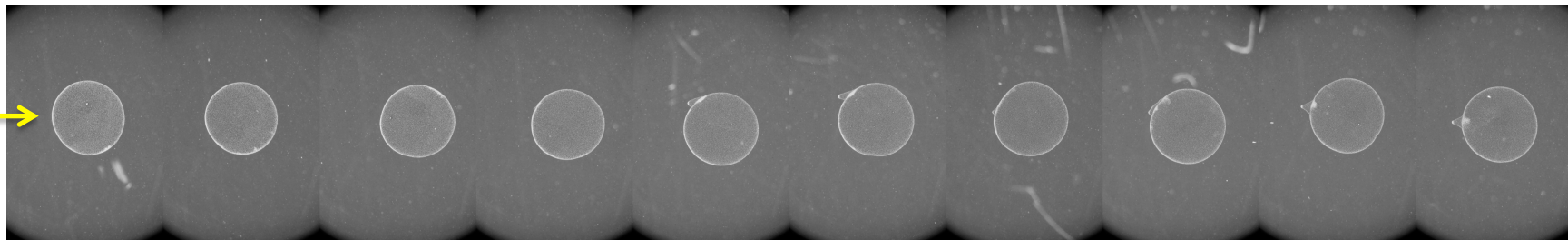
\*\* Some drops started with a heterogeneous distribution of cells (5<sup>th</sup>, 6<sup>th</sup>, 8<sup>th</sup>, 9<sup>th</sup>, 10<sup>th</sup> distances away from the LED), hence the extent of fingering from the cell-dense regions of these drops should not be considered when viewing the macroscopic data. The results from 19<sup>th</sup> Sept and 27<sup>th</sup> Sept can be used as a reference instead.



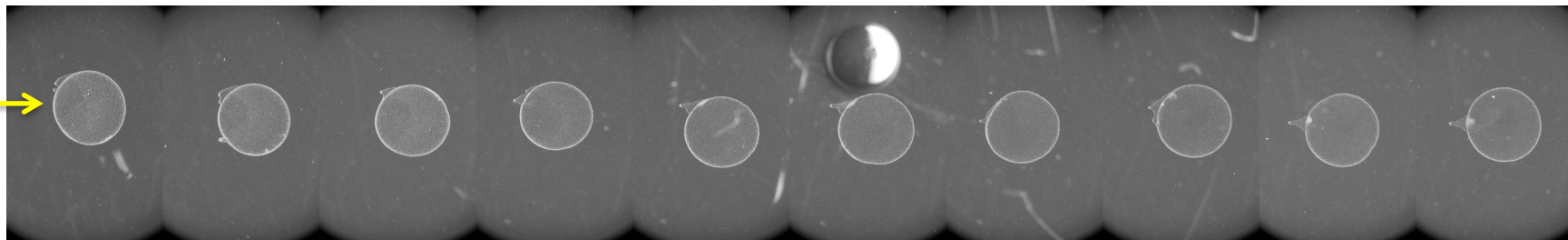
0hr



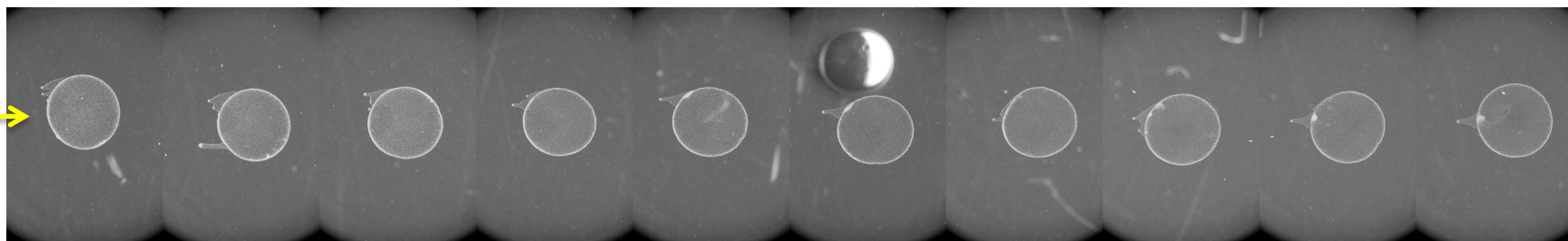
12hr



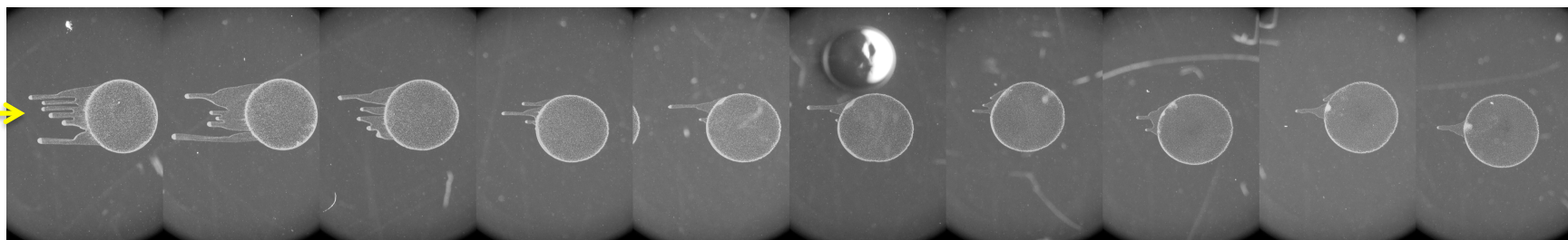
24hr

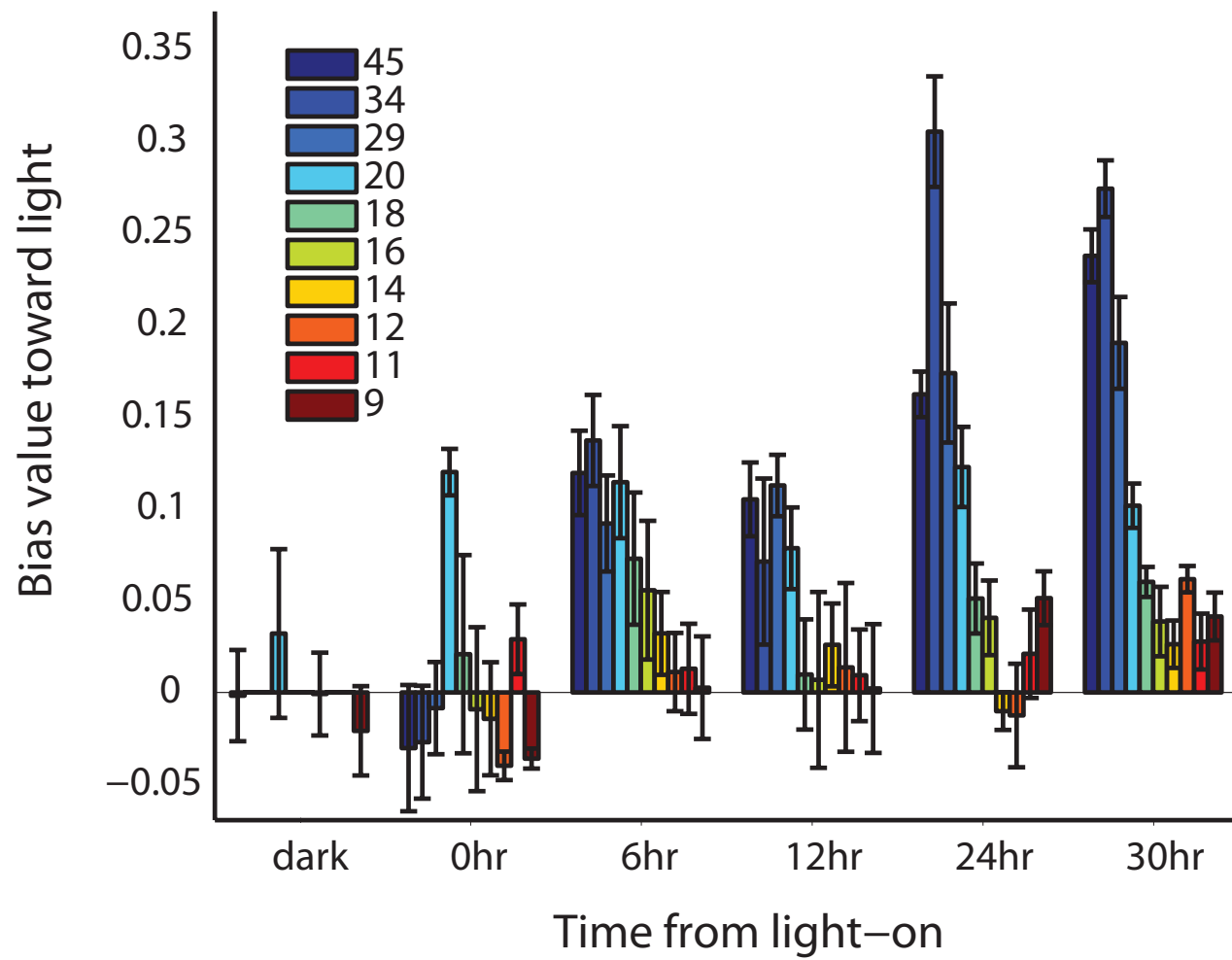


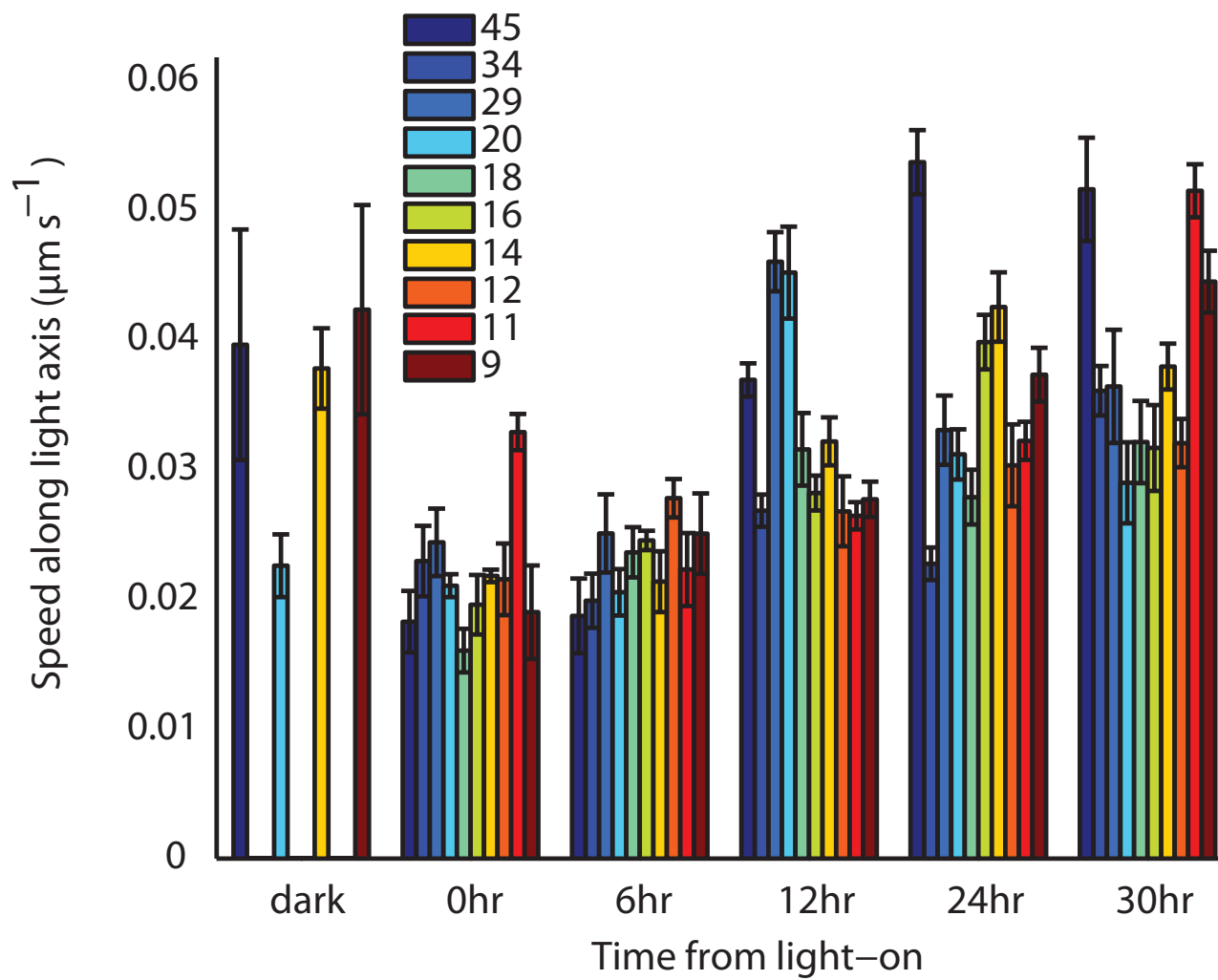
30hr



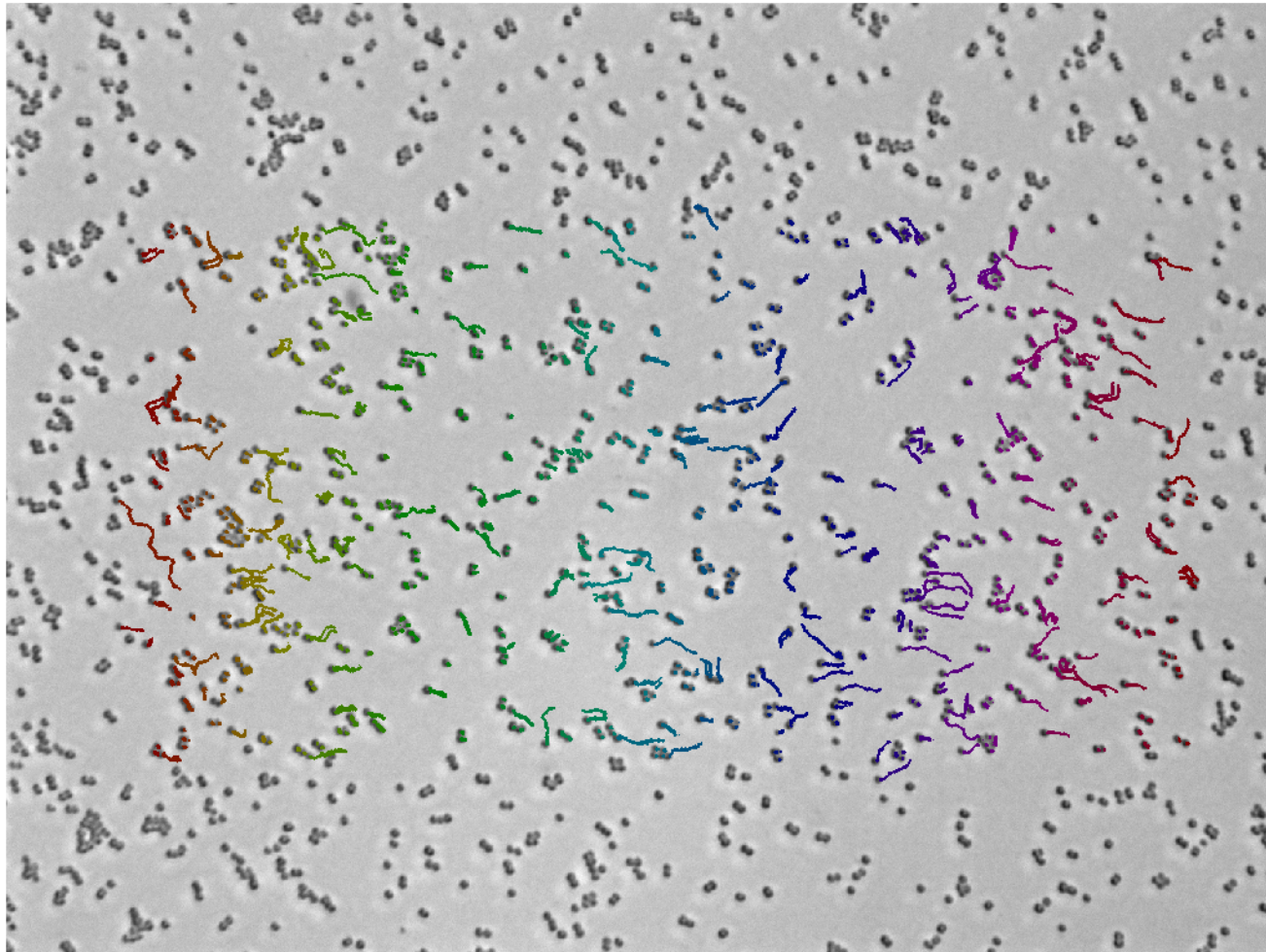
48hr





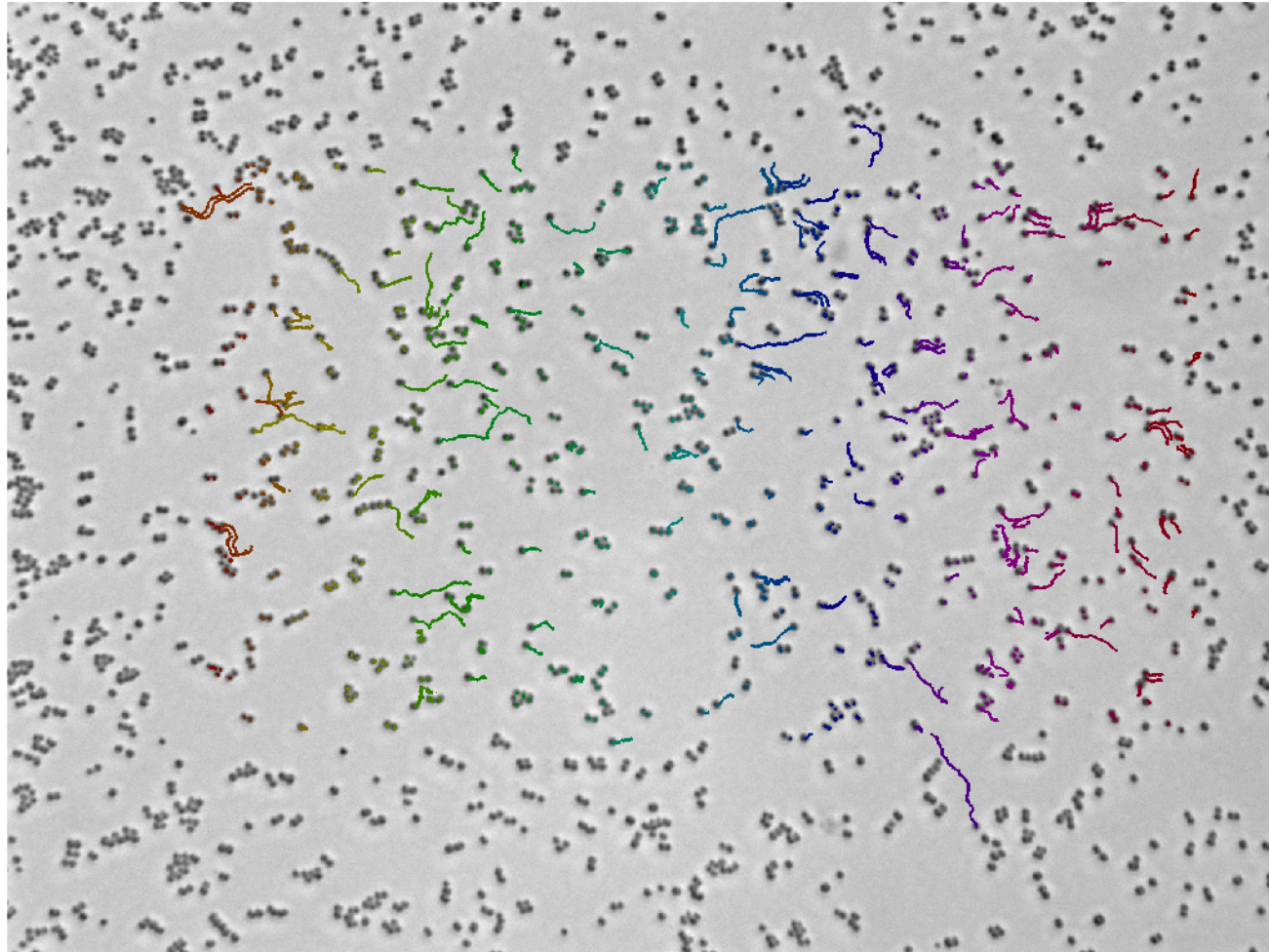


45uE @ 24 hrs

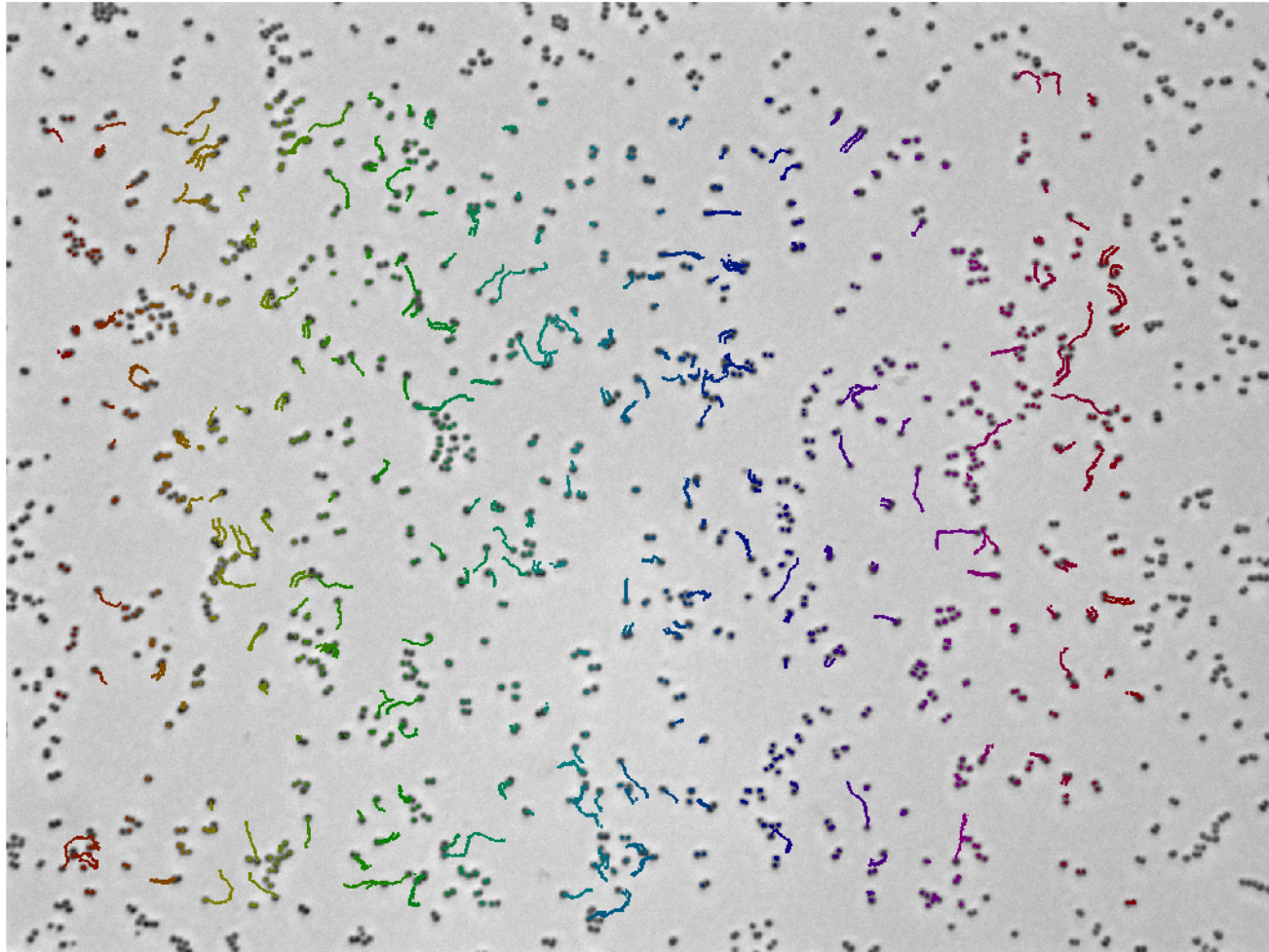




34uE @ 24 hrs



14uE @ 24 hrs



12uE @ 24 hrs

